

Cross sectional studies on the ectoparasites among rodents in scrub typhus cases in Karnal and Kaithal Districts of Haryana, India

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ABSTRACT: Orientia tsutsugamushi is a mite-borne bacterium belonging to the family Rickettsiaceae and is responsible for a disease called scrub typhus in humans, which is transmitted by the vector mite *Leptotrombidium deliense* (common ectoparasite on rodents) in most of Asian countries including India. The study conducted in selected villages of Karnal and Kaithal districts of Haryana state, India revealed four species of rodents - *Rattus rattus, R. norveigicus, Bandicota indica* and *Suncus murinus*. Dust mite *Dermatophagoides farina*; chigger mite *L. deliense and* fleas *Xenopsylla astia* and *X. cheopis* were prevalent on the rodents. © 2022 Association for Advancement of Entomology

KEY WORDS: Vector mite, chigger mite, dust mite, flea, rickettsial disease

Scrub typhus is a rickettsial disease caused by Orientia tsutsugamushi a mite-borne bacterium belonging to the family Rickettsiaceae. It is transmitted to humans by the bite of infected vector chigger mites. The trombiculid chigger mites are common ectoparasites on rodents and belong to the genus Leptotrombidium (Acariformes: Trombiculidae). Of these, the most common are L. pallidum (Nagayo), L. deliense (Walsh), L. scutellare (Nagayo) and L. akamushi (Brumpt) (Acosta-Jamett et al., 2020). Historically, scrub typhus had been endemic in Asia, Australia, and islands in the Indian and Pacific Oceans, known as the "tsutsugamushi triangle" (Bonell et al., 2017).

However, there have been recent reports of scrub typhus from Africa, the Middle East, and South America suggesting that the disease is no longer restricted to this triangle (Jiang and Richards, 2018). But no indigenous cases have been reported from North America and Europe. Scrub typhus is frequently reported from many Asian countries and is endemic in Nepal and its neighboring countries including India (Sub-Himalayan belt) and Bhutan, where it is considered an emerging infectious disease (Jeromie Wesley Vivian, 2017; Ranjan and Prakash, 2018; Tshokey *et al.*, 2018). Now it is re-emerging in almost in all states of India (Tilak and Kunte, 2019). The other vector mite spe-cies

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detected based on molecular techniques are Ascoschoengastia indica (Hirst), Guntheria cassiope (Womersley), Odontacarus sp. (Ewing), Eutrombicula wichmanni (Oudemans), and Microtrombicula chamlongi (Nadchatram and Kethley) (Elliott et al., 2019). These mites contribute for the transmission in different regions of India (Ranjan and Prakash, 2018). Knowledge of the vector, including species, distribution, density, and habitats, is important to understand the epidemiology of scrub typhus in a given area or region (Park et al., 2016). Clinical scrub typhus cases were reported in the districts of Karnal and Kaithal of Haryana state and most of them were confirmed in the laboratory of the PGIMER, Chandigarh. This study was undertaken with the main objective in finding out the vector mites among rodents that takes part in transmission of scrub typhus in the villages.

Study site: Haryana State has been contributing more number of scrub typhus cases from Karnal, Kaithal, and Panchkula districts with reference to the monthly periodical reports of National Centre for disease control (NCDC), Delhi, Based on that, a cross sectional entomological investigation was made in Sagger (29º41'8.2644"N; 76º59'25"E) and Dadapur (28ºN; 76.6629ºE) villages of Karnal District; Pundri (29.7621°N; 76.5546°E) and Harsola (29º48'5.51"N; 76º23'58.52"E) villages of Kaithal District of Haryana State. The GPS locations of areas from where rodents were trapped. Sherman and Wonder's traps were used. Single trap was placed in the scrub typhus positive house and two traps in the neighboring houses in each village. The sources of rat dwelling were identified by the presence of burrows, their paws, and excreta. Traps were laid at 6 PM and were collected in the next day morning by 6 AM (12 hours). A piece of Chapatti and roasted coconut were used as rodent baits. Rodents were identified using morphometric characteristics (Agrawal, 2000). Chigger mites were gathered from the rodents after they were euthanized using chloroform (Sigma-Aldrich, Bangalore, India) as described previously (Park et al., 2016). The chiggers were identified using the standard key for iden-tification of Indian *Trombiculidae* (Fernandes and Kulkarni, 2003). The identification features included the shape of the scutum, specialized leg setae, palpal chaetotaxy and chelicerae (Kuo *et al.*, 2015, Kumlert *et al.*, 2018; Philip *et al.*, 2021). The confirmation of the genus and species were based on pub-lished keys (Fernandes and Kulkarni 2003; Philip *et al.*, 2021). The dust mites were identified using the pictorial keys (Calloff and Stewart, 1997).

About 500g of soil and litter (humus samples) in and around rat burrows was collected and packed in zipped sachets and brought to the laboratory for detection of chigger mites employing Berlese's Funnel method. Berlese funnels are used for extracting the arthropods from soil and litter samples (Philip *et al.*, 2021). The organisms were identified using a binocular microscope (Dewinter) at 100X magnification. The Indices have been computed adopting the following standard formulae (Philip *et al.*, 2021; Basker *et al.*, 2022).

Chigger index: It is exclusively for Mite Borne Disease. It is measured by number of chiggers infested by a single host.

Prevalence rate of mites: Number of Hosts with ectoparasites / Total number of hosts examined

Mean intensity of ectoparasites from host animals: Total number of ectoparasites collected / Number of hosts infected with ectoparasites

Total flea index = Total number of fleas collected (regardless of species), divided by the total number of hosts examined.

Among the 12 traps laid at villages, viz., Sagger, Dadapur of Karnal District, and Pundri and Harsola villages of Kaithal District, six traps successfully trapped rodents. Four species of rats were encountered viz., *Rattus rattus, R. norveigicus, Bandicota indica and Suncus murinus*. Four species of ectoparasites isolated from the rats were - *Dermatophagaoides farinae, Leptotrombidium deliense* and fleas, *Xenopsylla astia* and *X. cheopis*. A total of 39 ectoparasites of all species could be collected. Among the mites, *D. farinae* was more prevalent (22 nos. and 56.4%) in rodents

Village	Rodents/ source	Ectoparasites - no./ species/(%)		
Sagger	Rattus rattus	1 Dermatophagoides spp (4.55%)		
Dadapur	Bandicota indica	17	17 Dermatophagoides spp (77.27%)	
		5	<i>Trombiculid</i> spp (33.3%)	
Dadapur	Suncus murinus	6	<i>Trombiculid</i> spp (40%)	
Pundri	R. rattus	1	female Xenopsylla cheopis	
		1	male X. astia	
		3	Trombiculidae spp (33.3%)	
		2	Dermatophagiodes spp (9.09%)	
Harsola	R. norveigicus	1	Trombiculidae spp (6.7%)	
		2	Dermatophagoides spp (9.09%)	
Dadapur	soil and litter	2	Dermatophagiodes spp.	

Table 1 Prevalence of ectoparasites among the rodent species

Table 2. Ectoparasites and their indices in scrub typhus reported villages in Karnal and Kaithal

Village (District)	Rodent	Dust mites – no./ (%)/ Index	Chiggers spp no./ (%)/Index	Fleas no./ Index	Prevale- nce rate	Mean intensity
Sagger (Karnal)	Rattus rattus	1 - (4.54%) - 0.5	0 - 0 - 0	0-0	0	0
Sagger (Karnal)	Suncus murinus	0	0 - 0 - 0	0-0	0	0
Dadapur (Karnal)	Bandicota indica	17 - (77.27%) - 17	5 - (33.3%) - 5	0-0	1	14
Dadapur (Karnal)	S. murinus	0	6 - (40%) - 6	0 - 0	1	14
Pundri (Kaithal)	R. rattus	2 - (9.09%) - 2	3 - (20%) - 3	2* - 1	1	7
Harsola (Kaithal)	Rattus norveigicus	2 - (9.09%) - 2	1 - (6.7%) -1	0	1	3

*1 female Xenopsylla cheopis and1 male X. astia flea

followed by chigger mite *L. deliense* (15 nos. and 38.4%) and species of fleas (2 and 5.1%). All the species were found on the rodents. From the soil and litter sample extraction *D. farinae* (2 no.) could be recorded (Table 1).

Chigger trombiculid mites, *L. deliense* were infested maximum on *S. murinus* (40%) followed by in *B. indica* (33.3%), *R. rattus* (20%) and *R. norveigicus* (6.7%). Dadapur of Karnal, was identified as more vulnerable for scrub typhus. *D.*

farinae was more detected in Dadapur and most of them were infesting *B. indica*. The soil sample collected in and around the rodent burrows from Dadapur village also showed the presence of D. farinae. The chigger index of Dadapur was 5.5; in Pundri, chigger index was 3 and in Horsala of Kaithal district, it is 2. Among these villages, Dadapur showed higher chigger index. Since these indices are greater than the critical chigger index of 0.69 (Olson et al., 1979; Basker et al., 2022), the probability of scrub typhus transmission in these villages is more. The prevalence rate of mite is one for all the rodent species. Mean intensity of ectoparasites from host animals is 14 each for *B*. indica and S. murinus. Of the fleas detected there were both the primary and secondary vectors of plague and were isolated from *R. rattus* which was captured in the semi-urban of Kaithal. Mean ectoparasites present per animal is dust mites 3.66, chigger mites 2.50 and fleas 0.33 (Table 2).

The bacteria *O. tsutsugamushi* cause scrub typhus and is transmitted by *Leptotrombidium* mites. It is responsible for a potentially fatal tropical infection which is a grossly under-recognized public health problem in India (Bonell *et al.*, 2017; Behera *et al.*, 2019). This disease is known to occur in diverse ecological settings in India with large numbers of cases being reported from Tamil Nadu, Andhra Pradesh, Karnataka and Kerala in the South, Himachal Pradesh, Uttaranchal, Jammu and Kashmir in the North, Meghalaya, Assam and Nagaland in the North-East, West Bengal and Bihar in the East and Maharashtra and Rajasthan in the West (Xu *et al.*, 2017; Philip *et al.*, 2021).

Dust mites feed mostly on dead skin and hairs shed from humans. In Haryana state, dust mites *Dermatophagoides pteronyssinus* and *Chyletus malaccenus* were previously reported (Voorhorst *et al.*, 1969). House dust mites, especially certain species of pyroglyphids, are the cause of allergic reactions of the respiratory tract (asthma and rhinitis). In recent years, house dust mite allergy has been identified as a frequent cause of asthma, especially among children (Traub and Wisseman Jr, 1968). Rodents are not only the reservoir of O. tsutsugamushi the causative organism for scrub typhus but also known to transmit human and animal diseases such as Leptospira spp, Borrelia spp., Yersinia pestis, and Bartonella spp. (Raharivolona and Ganzhone, 2009). In the present study, the potential primary and secondary plague vectors in India X. cheopis and X. astica have been isolated from the R. rattus which was captured from Pundri semi-urban of Kaithal district, Haryana and these species are potential enough to enhance the spread of plague within the communities in future if an outbreak occurs (Eisen et al., 2014). It is imperative that high priority be given to the research and development of effective integrated rodent management programs against domestic, peridomestic, and sylvatic rodent species to reduce the chances of parasite transmission. Since dust mite and chigger mite infestation found in rodents and soil, the surveillance on morbidity related to scrub typhus, allergy and other complications in the human community is highly essential.

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REFERENCES

Acosta-Jamett G., Martínez-Valdebenito C., Beltrami E., Silva-de La Fuente M.C., Jiang J., Richards A.L., Weitzel T. and Abarca K. (2020) Identification of trombiculid mites (Acari: Trombiculidae) on rodents from Chiloé Island and molecular evidence of infection with Orientia species. PLOS Neglected Tropical Diseases 14: e0007619.

- Agrawal V.C. (2000) Taxonomic studies on Indian Muridae and Hystricidae: Mammalia, Rodentia. Records of the Zoological Survey of India. Zoological Survey of India, Kolkatta, India.
- Basker P., Simmi T., Ajit S., Tushar N. and Sujit Kumar Singh (2022)Vector-Borne Zoonoses in India: Systematic gap analysis and Implications for Policy-level Interventions. Journal of Development Economics and Management Research Studies 09 (11): 96-126.
- Behera B., Biswal M., Das R.R., Dey A., Jena J. and Dhal S. (2019) Clinico-epidemiological analysis of scrub typhus in hospitalised patients presenting with acute undifferentiated febrile illness: A hospital-based study from Eastern India. Indian Journal of Medical Microbiology 37(2): 278–280. https://doi.org/10.4103.
- Bonell A, Lubell Y, Newton PN, Crump JA, Paris DH (2017) Estimating the burden of scrub typhus: a systematic review. PLOS Neglected Tropical Diseases11(9): e0005838.
- Colloff M. and Stewart G. (1997) House dust mites. In: Asthma. eds. Barnes P., Grunstein M.M., Leff A.R., Woolcock A., Philadelphia: Lippincott Raven. pp 1089-1104.
- Eisen R.J., MacMillan K., Atiku L.A., Mpanga J.T., Zielinski-Gutierrez E., Graham C.B., et al. (2014) Identification of Risk Factors for Plague in the West Nile Region of Uganda. American Journal of Tropical Medicine and Hygiene 90(6): 1047– 1058. doi: 10.4269/ajtmh.14-0035.
- Elliott I., Pearson I., Dahal P., Thomas N.V., Roberts T. and Newton P.N. (2019) Scrub typhus ecology: a systematic review of *Orientia* in vectors and hosts. Parasites and vectors 12(1): 513. https:// doi: 10.1186/s13071-019-3751-x
- Fernandes S. and Kulkarni S.M. (2003) Studies on the trombiculid mite fauna of India. Records of the Zoological Survey of India. Zoological Survey of India, Kolkata, India.
- Jeromie Wesley Vivian T., Ravi V., Leonard M., Govindakarnavar A., Samir V.S., Kamran Z., Tarun B., KSH S, Arun K., Jazeel A., *et al.* (2017) Risk factors for acquiring scrub typhus among children in Deoria and Gorakhpur Districts, Uttar Pradesh, India. Emerging infectious diseases Journal 24(12): 2364.

- Jiang J and Richards AL (2018). Scrub typhus: no longer restricted to the Tsutsugamushi triangle. Tropical Medicine and Infectious Disease 3(1):11-13.
- Kumlert R., Chaisiri K., Anantatat T., Stekolnikov A.A., Morand S., Prasartvit A., Makepeace B.L., Sungvornyothin S. and Paris D.H. (2018) Autofluorescence microscopy for pairedmatched morphological and molecular identification of individual chigger mites (Acari: Trombiculidae), the vectors of scrub typhus. PLoS One 13: e0193163.
- Kuo C.C., Lee P.L., Chen C.H. and Wang H.C. (2015) Surveillance of potential hosts and vectors of scrub typhus in Taiwan. Parasites & Vectors 8: 611.
- Olson J.G. (1979) Forecasting the onset of a scrub typhuss epidemic in the Pescoders Islands of Taiwan using daily maximum temperatures. Tropical and geographical medicine 31: 519-524.
- Park J.W., Chung J.K., Kim S.H., Cho S.J., Ha Y.D., Jung S.H., Park H.J., Song H.J., Lee J.Y., Kim D.M., et al. (2016) Sero-epidemiological survey of zoonotic diseases in small mammals with PCR detection of *Orientia tsutsugamushi* in chiggers, Gwangju, Korea. Korean Journal of Parasitology 54: 307– 313.
- Philip S.P., Renu G., Ranganathan K. and Jaganathasamy N. (2021) Ectoparasites of some wild rodents / shrews captured from Scrub typhus reported areas in Tamil Nadu, India, International Journal of Acarology 47(3): 218-221. https://doi.org/ 10.1080/01647954.2021.1887932.
- Raharivolona B.M. and Ganzhone J.U. (2009) Gastrointestinal parasite infection of the gray mouse lemur (*Microcebus murinus*) in the littoral forest of Mandana Madagascar: effects of forest fragmentation and degradation. Madagascar conservation & development 4(2): 103-112. doi:10,4314/mcd.v4i2.48650.
- Ranjan J. and Prakash J.A.J. (2018) Scrub typhus reemergence in India: contributing factors and way forward. Medical Hypotheses 115: 61–64. DOI: 10.1016/j.mehy.2018.03.019.
- Tilak R. and Kunte R. (2019) Scrub typhus strikes back: are we ready? Medical Journal Armed Forces India 75: 8-17.
- Traub R. and Wisseman Jr. C.L. (1968) Ecological considerations in scrub typhus-species. Bulletin of the World Health Organization 39(2): 219-230.
- Tshokey T., Stenos J., Durrheim D.N., Eastwood K., Nguyen C., Vincent G. and Graves S.R. (2018)

Rickettsial Infections and Q Fever Amongst Febrile Patients in Bhutan. Tropical Medicine and Infectious Disease 3(1): 12. doi.org/10.3390/ tropicalmed301 0012.

Voorhorst R., Spieksma F.T.M. and Varekamp H. (1969) House dust atopy and the house dust mite *Dermatophagoides pteronyssinus* (Trouessart 1897). Leiden, The Netherlands, Stafleu's Scientific Publishing Company.

Xu G., Walker D.H., Jupiter D., Melby P.C. and Arcari C.M. (2017) A review of the global epidemiology of scrub typhus. PLOS Neglected Tropical Diseases 11(11): e0006062. https: // doi.org/10.1371.

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