



## Genitalia study on the genus *Glyphodes* (Crambidae: Spilomelinae) in Tamil Nadu, India

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**ABSTRACT:** Survey carried out to collect species belonging to the genus *Glyphodes* in different places of Tamil Nadu, viz., Coimbatore, Anaikatti, Ooty, Yercaud, Kodaikanal and Periyakulam recorded a total of six species of the genus *Glyphodes* viz., *G. bivitalis*, *G. caesalis*, *G. canthusalis*, *G. onychinalis*, *G. pulverulentalis* and *G. stolalis*. Male and female genitalia of the collected species are described. © 2017 Association for Advancement of Entomology

**KEYWORDS:** Distribution, *Glyphodes*, Spilomelinae, genitalia, Tamil Nadu

### INTRODUCTION

Spilomelinae is the largest subfamily in the Crambidae, with about 3,767 species worldwide (Regier *et al.*, 2012) and are of economic importance as many species cause serious damage to agricultural and horticultural crops, forests trees and ornamental plants (Mathew and Menon, 1984). Under Spilomelinae the genus *Glyphodes* Guenée consists of 120 species and is widespread in tropical regions, with some species penetrating into subtropical and warm temperate areas (Common, 1990; Robinson *et al.*, 1994). Twenty five species have been recorded in the Southeast Asia and 17 species in Australia (Robinson *et al.*, 1994; Shaffer *et al.*, 1996). In Tamil Nadu, three species were recorded by Fletcher (1914) and Nair (1970) are *Glyphodes bivitalis*, *G. caesalis*, *G. canthusalis*. Before this background, the present study aims to identify the *Glyphodes* species encountered in different regions of Tamil Nadu by studying morphological characters of the genitalia, and to record the distribution of those species.

### MATERIALS AND METHODS

The study was conducted in Coimbatore, Anaikatti, Ooty, Yercaud, Kodaikanal and Periyakulam areas of Tamil Nadu, India during 2014-15. Moths were attracted using a white cloth (1.5 x 5.5 m) and a mercury lamp (400 Watts) from 6.00pm to 6.00am. The moths were killed with ethyl acetate and transferred into butter paper covers. The insects were curated and labeled as per Johnson and Triplehorn (2005). The moths were identified as per Hampson (1896, 1898) and with the reference collection of the Insect Biosystematics Laboratory, TNAU, Coimbatore. The generic and species nomenclature followed as per Beccaloni *et al.* (2003) and Nuss *et al.* (2003-2015). Taxonomically informative characters viz., antennae, labial palpi, forewing (FW), hindwing (HW), legs, tympanum, male and female genitalia were studied. The standard technique given by Robinson (1976) was followed for genitalia studies, while wing venation was studied using the Comstock – Needham system (1898). Genitalia images were taken in a

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Leica MZ16 stereozoom microscope. Illustrations were drawn by using mirror type camera lucida Leica M80.

## RESULTS AND DISCUSSION

During the study, six species of the genus *Glyphodes* were collected from light trap: *Glyphodes bivitalis* (Yercaud, Coimbatore), *G. caesalis* (Anaikatti, Yercaud), *G. canthusalis* (Coimbatore), *G. onychinalis* (Coimbatore, Yercaud, Kodaikanal), *G. stolalis* (Coimbatore, Yercaud, Kodaikanal), *G. pulverulentalis* (Coimbatore, Killikulam, Kodaikanal, Periyakulam); the later was also recorded from its host plant (mulberry).

Common characters of the six investigated species: Filiform antennae; labial palpi upturned; maxillary palpi and basally scaled proboscis present. In the forewing, vein  $R_3$  stalked with  $R_4$ ;  $R_5$  free; 1A+2A complete with a basal fork and 3A short, forming a loop. In hind wing,  $Sc+R_1$  thickened, basally anastomosed, with  $R_s$  after its origin from upper angle of cell and ends at costa near apex. The male and female genitalia of six species are described below.

### *Glyphodes bivitalis* Guenée, 1854

**Male genitalia:** Uncus long, anteriorly enlarged with constricted tip; uncus tip with five short spines, ventrally fringed with long hairs. Gnathos absent; subscaphium long, strap-like. Tegumen long, sclerotized, dome-shaped and arched; vinculum short; saccus broadly U-shaped. Valva long, membranous; apex broadly fringed with hairs; costa inconspicuous; medially sclerotized line with distinct thickening; sacculus broad at base, ridge-like, dorsally bent inward; harpe long, claw-like basally supported with medial sclerotized line; hair pencil with bunch of short hairs. Transtilla composed of triangular anterior projections meeting at mid line; juxta narrow, flap-like. Phallus long, sclerotized thread-like (Fig. 1a).

**Female genitalia:** Anal papillae densely setose. Anterior apophyses long, basally with angular

projection; posterior apophyses short when compared to anterior apophyses. Ostium oval, membranous; antrum short, moderately sclerotized; ductus seminalis originate at posterior end of ductus bursae above antrum. Ductus bursae very long, narrow and membranous; corpus bursae oval, membranous; signum composed of two denticulate, sclerotized signa (Fig. 2a)

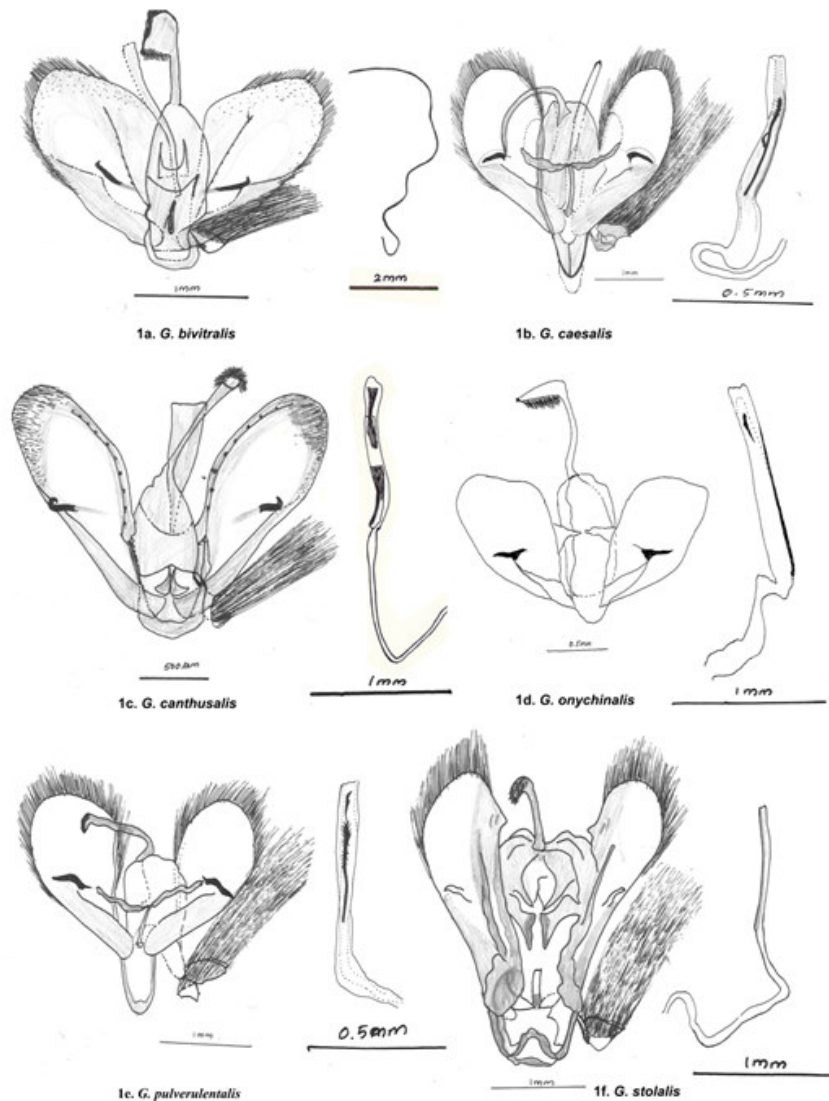
### *Glyphodes caesalis* Walker, 1859

**Male genitalia:** Uncus long and narrow, apex enlarged and pointed, tip fringed with hairs. Gnathos absent; subscaphium long, strap-like. Tegumen longer than wide, sclerotized and arched; vinculum long, sclerotized and saccus long, U shaped. Valva long, membranous, apex narrow and rounded; costa inconspicuous, dorsally fringed with long hairs; sacculus broad, weakly sclerotized; ridge like dorsal edge with fine setae; harpe long, spine-like, sclerotized. Transtilla long, sclerotized band-like meeting at mid line; juxta narrow, sclerotized, arrow-like. Phallus almost straight, vesica with long sclerotized bar with lateral spine-like projection; medially long, curved sclerotized hook-like cornuti (Fig. 1b).

**Female genitalia:** Anal papillae fringed with both long and short setae. Anterior apophyses long, basally with angular projection; posterior apophyses short. Ostium oval, membranous. Antrum short, sclerotized; ductus seminalis originate at posterior end of ductus bursae below antrum. Ductus bursae long and wide, membranous; caudally constricted; corpus bursae oval, membranous; sclerotized dots forming two strip-like signa (Fig. 2b).

### *Glyphodes canthusalis* Walker, 1859

**Male genitalia:** Uncus long, narrow, anterior tip enlarged and round, dorsally fringed with long setae. Gnathos absent, subscaphium long, strap-like, uniformly sclerotized. Tegumen long, membranous; vinculum long, curved and saccus broadly U shaped. Valva long, medially widened and apex narrow fringed with hairs; costa sclerotized, prominent with series of spots; sacculus broad at base, narrowed anteriorly and prominent; harpe long, sclerotized,

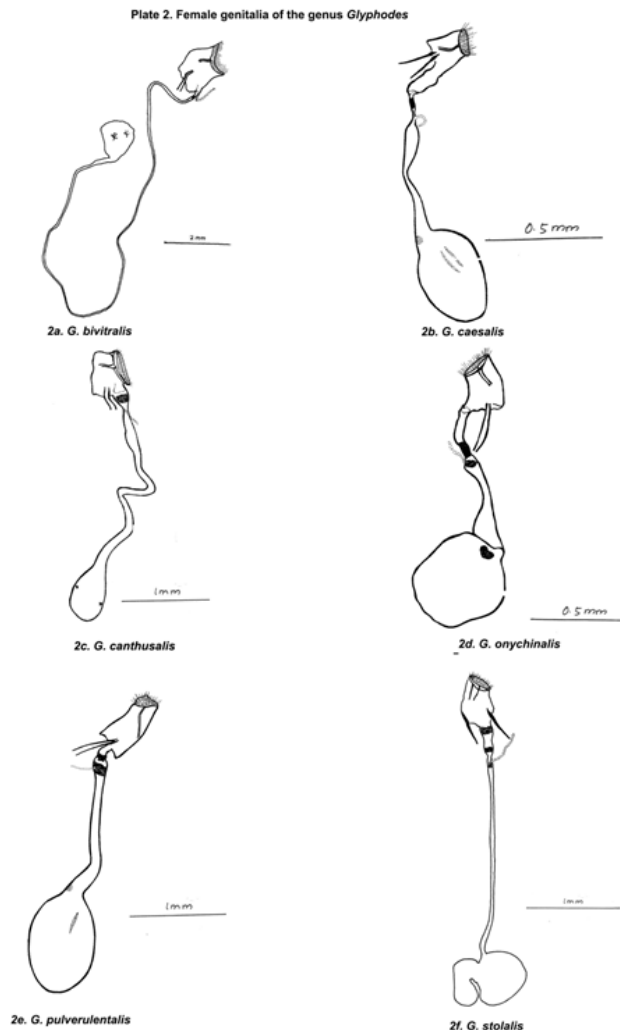
Plate 1. Male genitalia of the genus *Glyphodes*

claw-like; upturned apically. Transtilla membranous; juxta broad with anterior weakly sclerotized projection. Phallus long, vesica with intermittent patch-like sclerotization (Fig. 1c).

**Female genitalia:** Anal papillae densely fringed with setae. Anterior apophyses longer than posterior apophyses with medial angular projection. Ostium funnel shaped, membranous; ductus seminalis originate at posterior end of ductus bursae. Ductus bursae long, membranous; corpus bursae elongate, membranous; two small patches of sclerotized spines forming signa (Fig. 2c).

#### *Glyphodes onychinalis* (Guenée, 1854)

**Male genitalia:** Uncus long, anterior half strongly curved, enlarged and beak shaped, ventrally fringed with long hairs; anterior tip with four short spines. Gnathos absent; subscaphium long, strap-like. Tegumen longer than wide, arched; vinculum almost long, weakly sclerotized and saccus broadly U shaped. Valva long, apex narrow; outer margin covered with black hairs; costa inconspicuous; sacculus narrow, ridge-like, weakly sclerotized; harpe long, spine-like; directed distally and supported basally by the apical tip of the



sacculus. Transtilla composed of lateral projections meeting mid line; juxta narrow, flap-like and weakly sclerotized. Phallus almost straight, strongly sclerotized on ventral side and vesica with sclerotized patch-like cornutus (Fig. 1d).

**Female genitalia:** Anal papillae densely setose. Anterior apophyses long, basally with angular projection; posterior apophyses short. Ostium oval, membranous. Antrum broadly sclerotized; ductus seminalis originate at antrum. Ductus bursae long, wide at anterior and membranous; corpus bursae round, membranous with heart shaped signum (Fig. 2d).

### *Glyphodes pulverulentalis* Hampson, 1896

**Male genitalia:** Uncus long and narrow, anterior tip enlarged and pointed, beak-shaped dorsally with short setae. Gnathos absent; subscaphium long, strap-like. Tegumen longer than wide, sclerotized and arched; vinculum long, sclerotized and saccus long, U shaped. Valva long, membranous, apex broadly rounded; costa weakly sclerotized, dorsally fringed with long hairs; sacculus broad, weakly sclerotized; ridge-like, dorsal edge with fine setae; harpe long, spine-like; sclerotized, directed distally. Transtilla long, sclerotized band-like meeting at mid line; juxta narrow, sclerotized, arrow-like. Hair pencil with long bunch of hairs; phallus almost straight, vesica with long sclerotized bar with lateral spine-like projection; apically with long, curved sclerotized hook-like cornutus (Fig. 1e).

**Female genitalia:** Anal papillae oval, fringed with both long and short setae. Anterior apophyses long, basally with angular projection; posterior apophyses short. Ostium membranous; antrum broadly sclerotized; ductus seminalis originate from antrum. Ductus bursae long, membranous; corpus bursae oval, membranous; signum as two strip-like sclerotized dots (Fig. 2e).

### *Glyphodes stotalis* Guenée, 1854

**Male genitalia:** Uncus long and narrow, anterior tip enlarged and spoon shaped, fringed with setae. Gnathos absent; tegumen longer than wide, arched and teguminal ridges strongly sclerotized; vinculum almost long, sclerotized and saccus broadly W shaped. Valva long, narrow at base with broad apex, outer margin fringed with long hairs; costa inconspicuous, sclerotized, distally with triangular dorsal projection; sacculus narrow, ridge-like; harpe long, sclerotized and spine-like. Transtilla sclerotized flap-like extending downward; juxta short, membranous. Hair pencil with short bunch of hairs. Phallus straight, long; phallus apodeme strongly sclerotized on ventral side; cornuti absent (Fig. 1f).

**Female genitalia:** Anal papillae oval, fringed with short and long setae. Anterior apophyses long,

**Plate 3. Identified species of the genus *Glyphodes***



**3a. *G. bivitalis***



**3b. *G. caesalis***



**3c. *G. canthusalis***



**3d. *G. onychinalis***



**3e. *G. pulverulentalis***



**3f. *G. stolalis***

basally broad; posterior apophyses short. Ostium oval, weakly sclerotized; antrum weakly sclerotized; ductus seminalis originate at posterior end of ductus bursae below antrum. Ductus bursae long, narrow; corpus bursae rounded, membranous; accessory bursae present, dropper shaped with minute denticulation; signum absent (Fig. 2f).

### Identification key to investigated species of *Glyphodes*

1. FW and HW with spots and striations; ♀ genitalia with signum, appendix bursae absent ..... 2
- 1a. FW and HW with striations; ♀ genitalia without signum, appendix bursae present ..... *Glyphodes stolalis*
2. ♀ genitalia with denticulate or stripe-like signum ..... 3
- 2a. Heart shaped signum..... *Glyphodes onychinalis*
3. Uncus long and narrow; tip enlarged ..... 4
- 3a. Uncus beak shaped; tip with five short spines ..... *Glyphodes bivitalis*
4. Costa with series of dots; cornuti absent ..... *Glyphodes canthusalis*
- 4a. Costa without dots; cornuti present
5. Ductus bursae uniform tube-like; ductus seminalis originates at antrum .....  
.....*Glyphodes pulverulentalis*
- 5a. Ductus bursae constricted and enlarged; ductus seminalis originate below antrum .....  
..... *Glyphodes caesalis*

Hampson (1896) described 48 species in the genus *Glyphodes* in the Fauna of India. Many species originally described in *Glyphodes* are currently referable under several genera such as *Palpita* Hübner, 1808, *Parotis* Hübner, 1831, *Stemorrhages* Lederer, 1863, *Arthroschista* Hampson, 1893 (Mathew, 2006). Sutrisno (2002, 2003) studied the

phylogenetic relationship among the Australian *Glyphodes* group and 17 genera which are morphologically similar. According to Sutrisno, the genus *Glyphodes* is not a monophyletic group and the genitalia characters of *G. bivitalis* and *G. stolalis* are also confirmed with his study. *Glyphodes pulverulentalis* is a serious pest of mulberry in Karnataka, Andhra Pradesh and Tamil Nadu (Geetha Bai *et al.*, 1997; Samuthiravelu *et al.*, 2010; Rahmathulla *et al.*, 2011). The pest is also reported in different locations of India *viz.*, Nagaland (Gupta, 1994), Jammu (Sharma and Tara, 1985), Kashmir (Dar, 1993) and Punjab (Mavi *et al.*, 1996). Fletcher (1914) reported *G. caesalis* as a pest of jackfruit in Karnataka and Maharashtra. It is also recorded from Assam, Sikkim, Bihar, Uttar Pradesh, Andhra Pradesh and Tamil Nadu of India (Chowdhury and Majid, 1954; Prarthna *et al.*, 2014). Soumya *et al.* (2015) reported *G. caesalis* for the first time from Kerala. In India, Kirti and Sodhi (2001) recorded five species (*G. bicolor*, *G. caesalis*, *G. canthusalis*, *G. pulverulentalis*, *G. stolalis* and *G. zelleri*) from North-Eastern India, Rose (2001) recorded seven species (*G. actorionalis*, *G. caesalis*, *G. multilinealis*, *G. canthusalis*, *G. pulverulentalis*, *G. stolalis* and *G. zelleri*) from Assam.

Tamil Nadu is endowed with rich flora and fauna that contribute to the biodiversity. Despite *Glyphodes* being a species rich tropical genus. Only six species were recorded during the present study. The diversity of Crambidae may vary in different locations with different climatic conditions. Therefore proper collection, identification and documentation of these species provide the most reliable data for conservation and management practices.

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### REFERENCES

- Beccaloni G., Scoble M., Kitching I., Simonsen T., Robinson G., Pitkin B., Hine A. and Lyal C. (2003).

- The Global Lepidoptera Names Index (LepIndex). <http://www.nhm.ac.uk/entomology/lepindex>
- Bethuene-Baker G.T. (1914). Notes on the taxonomic value of genital armature in Lepidoptera. Trans. Entomological Society, London, 314-337.
- Chowdhuri S and Majid S. (1954) Hand book of plant protection. Department of Agriculture, Assam, Shillong. 117 pp.
- Common I.F.B. (1990). Superfamily Pyraloidea. In: Moths of Australia, Melbourne University Press, Carlton. 343-358, pp.
- Comstock J.H. and Needham J.G. (1898) The wings of Insects. Chapter III. The specialization of wings by reduction. The American Naturalist 32: 231-257.
- Dar M. A. (1993). Observation the biology of *Glyphodes pyloalis* Walker (Pyralidae: Lepidoptera) a serious pest of mulberry. The collection of papers of the II International Silk Conference, China, pp. 1-4.
- Eyer J.R. (1926) The morphological significance of the juxta in the male genitalia of Lepidoptera. Bulletin of Brooklyn Entomological Society 32-37.
- Fletcher T. B. (1914) Some South Indian Insects. Indian Agricultural Research Institute, New Delhi, India. 565pp.
- Geetha Bai M. and Marimadaiah B. (2002) Seasonal occurrence of the mulberry leaf-roller *Diaphania pulverulentalis* (Hampson) and its parasitoids. Entomol 27(1): 51-56.
- Gupta S. L. (1994) Check list of Indian Pyraustinae. (Lepidoptera :Pyralidae). Memoirs of the Entomological Society of India 14, 1-87.
- Hampson G.F. (1896) The Fauna of British India including Ceylon and Burma, Moths Vol. IV. Taylor and Francis Limited, London. 594 pp.
- Hampson G.F. (1898) A revision of moths of sub-family Pyraustinae and family Pyralidae. Part 2. Proceedings of Zoological Society of London. pp 590-761.
- Johnson N.F. and Triplehorn C.A. (2005) Borror and Delong's Introduction to the Study of Insects. Thompson-Brooks/Cole. California. 888pp.
- Kirti J.S. and Sodhi J.S. (2001) A systematic list of Pyraustinae of Northeastern India (Pyralidae: Lepidoptera). Zoo's print Journal 16(10): 607-614.
- Mathew G. (2006) An inventory of Indian pyralids (Lepidoptera: Pyralidae). Zoo's Print Journal 21(5): 2245-2258
- Mathew G. and Menon M.G.R. (1984) The pyralid fauna (Lepidoptera: Pyraloidea: Pyralidina) of Kerala (India). Journal of Entomological Research 8(1): 5-13.
- Mavi G. S., Bhalla J. S. and Mann A. S. (1996) *Glyphodes pyloalis* Walker (Pyralidae: Lepidoptera) pest of mulberry-a new record in Punjab. Journal of Sericulture 4(1), 28-29.
- Nair M. R. G. K. (1970) Insects and Mites of Crops in India. New Jack Printing Works Private Ltd, Bombay. 404pp.
- Niculescu E.V. (1969) Some aspects of the study of the genital armature in Lepidoptera. Studii Cerc. Biol. (Ser. Zool.) 21(2): 137-140.
- Nuss M., Landry B., Mally R., Vegliante F., Tränkner A., Bauer F., Hayden J., Segerer A., Schouten R., Li H., Trofimova T., Solis M. A., De Prins J. and Speidel W. (2003-2015) Global Information System on Pyraloidea. [www.pyraloidea.org](http://www.pyraloidea.org).
- Prarthna R., Debanand D., Baruah K. and Dutta S.K. (2014) New record of *Glyphodes caesalis* Walker (Lepidoptera: Pyralidae) as a pest of jackfruit from Jorhat (Assam), Insect Environment 19(4): 247.
- Rahmathulla V. K., Kishor Kumar C.M., Angadi B. S. and Sivaprasad V. (2011) Association of climatic factors on population dynamics of leaf roller, *Diaphania pulverulentalis* Hampson (Lepidoptera: Pyralidae) in mulberry plantations of sericulture seed farm. Psyche 2012: 1-6. doi:10.1155/2012/186214.
- Regier C.J., Mitter C., Solis M.A., Hayden J.E., Landry B., Nuss M., Simonsen T.J., Yen S.H., Zwick A. and Cummings M.P. (2012). A molecular phylogeny for the pyraloid moths (Lepidoptera: Pyraloidea) and its implications for higher-level classification. Systematic Entomology 37: 635-656.
- Robinson G. S. (1976) The preparation of slides of Lepidoptera genitalia with special reference to microlepidoptera. Entomological Gazette 27(2): 127-132.
- Robinson G. S., Tuck K. R. and Shaffer M. (1994) Field Guide to the Smaller Moths of Southeast Asia. Malaysian Nature Society, Kuala Lumpur.
- Rose H.S. (2001) An inventory of the moth fauna (Lepidoptera) of Jatinga, Assam, India. Zoos print journal 17(2): 707-721.
- Samuthiravelu P., Ravikumar J., Qadri S.M.H., Hemanthkumar L. and Jayaraj S. (2010) Influence of abiotic factors on population dynamics of leaf webber *Diaphania pulverulentalis* and its natural enemies in mulberry. Journal of Biopesticides 3(1): 037-042.
- Shaffer M. A., Nielsen E. S. and Horak M. (1996) Pyraloidea. In: Checklist of the Lepidoptera of Australia (Eds. Nielsen E. S., Edwards E. S. and Rangsit T. V.), CSIRO, Australia, 164-199 pp.

- Sharma B. and Tara J. S. (1985) Insect pests of mulberry plants (*Morus* sp.) in Jammu region of Jammu and Kashmir state. *Indian Journal of Sericulture* 24: 7-11.
- Soumya K., Krishnamoorthy K. A. and Venkatesha M. G. (2015) Occurrence of Jack shoot and fruit borer, *Diaphania caesalis* (Walker) (Pyralidae: Lepidoptera) in Kerala, India. *Current Biotica* 9(3):295-299.
- Sutrisno H. (2002) A preliminary study on relationships among selected Australian members of the tribe Spilomelini (Lepidoptera: Crambidae: Pyraustinae). *Zoological Science* 19: 915-929.
- Sutrisno H. (2003) Phylogeny of *Glyphodes* Guenee (Lepidoptera: Crambidae: Spilomelinae) based on nucleotide sequence variation in a mitochondrial CO I gene: congruence with morphological data. *Treubia* 33(1): 35-42.

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